

Flow Cytometry (FCM)

I. Preparation of Solutions and Reagents

- 1. PBS Buffer: Prepared using PBS powder and ddH2O. Cool to 4°C before use.
- 2. Fixative (optional): 4% Paraformaldehyde.
- 3. Permeabilization Solution (optional): 0.1% Triton X-100, prepared with Triton X-100 and PBS buffer. Store at 4°C.
- 4. Blocking Solution: 5% BSA, prepared with BSA and PBS buffer. Prepare fresh before use and store at 4°C.

II. Experimental Procedure

A. Sample Preparation

Cell Collection

- (1) Suspension cells: Centrifuge the cell culture medium directly and collect the cells.
- (2) Adherent cells: Discard the medium, wash once with PBS, digest the cells with trypsin, and collect the cells.
- (3) Tissue samples: Mince the tissue and digest with collagenase/trypsin, filter through a cell strainer to remove tissue clumps, and collect the cells.

Cell Counting and Suspension Preparation

- (1) Resuspend cells in 1 mL PBS, centrifuge at 1500 rpm for 5 minutes, discard the supernatant, and repeat the wash step 1–2 times.
- (2) Take a small amount of cell suspension, stain with trypan blue for viability assessment, and adjust the cell concentration to 1×106–5×106 cells/mL.

B. Cell Surface Antigen Staining (Permeabilization Not Required)

- 1. Blocking: Resuspend 1×10^6 cells in 300 μ L of blocking buffer. Incubate at room temperature for 15 minutes and protect from light. Centrifuge at 1500 rpm for 5 minutes, discard the supernatant, and wash the cells once with PBS buffer.
- 2. Antibody Incubation: Add an appropriate amount of fluorophore-conjugated antibody and mix gently. Incubate at 4°C for 20–30 minutes in the dark. (Refer to the product website for recommended antibody dilutions.)
- 3. Washing: Add 1 mL of PBS, centrifuge at 1500 rpm for 5 minutes, discard the supernatant, and repeat once.
- 4. Resuspension: Resuspend the cells in 500 μ L of PBS buffer. Filter through a 35–40 μ m cell strainer to remove aggregates, then transfer the cells to a flow cytometry tube.

C. Intracellular Antigen Staining (Membrane Permeabilization Required)

- 1. Fixation: Take 1×10⁶ cells and add 1 mL of 4% paraformaldehyde. Incubate at room temperature in the dark for 15-20 minutes. Centrifuge at 1500 rpm for 5 minutes, discard the supernatant, and wash the cells twice with PBS buffer.
- 2. Permeabilization: Add 500 μ L of 0.1% Triton X-100, vortex gently to mix, and incubate on ice in the dark for 10-20 minutes. Centrifuge at 1500 rpm for 5 minutes, discard the supernatant, and wash the cells once with PBS buffer containing 0.1% Triton X-100.
- 3. Blocking: Resuspend the cells in 300 μ L of blocking buffer and incubate at room temperature in the dark for 20 minutes. Centrifuge at 1500 rpm for 5 minutes, discard the supernatant, and wash the cells once with PBS buffer containing 0.1% Triton X-100.
- 4. Antibody Incubation: Add an appropriate amount of fluorescently labeled antibody, gently flick to mix, and incubate at 4 °C in the dark for 30-60 minutes (refer to our website for antibody dilution).
- 5. Washing: Resuspend the cells in 1 mL of PBS buffer containing 0.1% Triton X-100, centrifuge at 1500 rpm for 5 minutes, and discard the supernatant. Repeat the wash once with PBS buffer containing 0.1% Triton X-100, followed by one final wash with PBS buffer.



6. Resuspension: Resuspend the cells in 500 μL of PBS buffer. Filter through a 35–40 μm cell strainer to remove cell clumps, and transfer the cell suspension to a flow cytometry tube.

D. Flow Cytometry Analysis

Set the parameters for the flow cytometer and perform the analysis.

III. Experimental Notes

- 1. When using fluorescently labeled primary antibodies, secondary antibodies are not required. However, if the primary antibody is unlabeled, a fluorescently labeled secondary antibody (specific to the host species of the primary antibody) must be used.
- 2. For cell permeabilization, either 0.1% Triton X-100 or 90% methanol can be used. Triton X-100 is suitable for most intracellular antigens, while methanol provides more thorough permeabilization but may disrupt certain antigen conformations. It is recommended to perform a preliminary experiment to optimize conditions.

IV. Recommended Experimental Products

1. Fluorescently Labeled Antibodies for Flow Cytometry:

https://www.targetmol.com/antibodies/flow cytometry antibodies

Bovine Serum Albumin (BSA), T5664

Triton X-100, T64297

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